## LETTER TO THE EDITOR

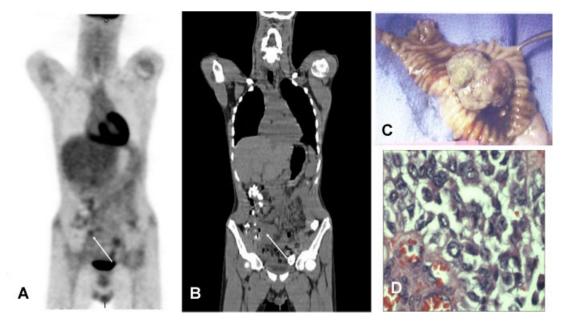
## Clear Cell Sarcoma of the Gastrointestinal Tract Presenting as a Second Malignant Neoplasm Following Neuroblastoma in Infancy

To the Editor: Clear cell sarcoma of the gastrointestinal tract (GI CSS) in children is extremely rare with only four cases reported in the literature [1–4]. Two of these reports describe CCS presenting as a second malignant neoplasm (SMN) following prior therapy for acute leukemia [1,2]. We are reporting the first published case of GI CCS presenting as an SMN from a survivor of neuroblastoma in infancy.

A 15-year-old Caucasian male presented with fevers, myalgias, night sweats, and weight loss. His past medical history was significant for neuroblastoma diagnosed at 2 months of age with disease localized to the liver. The tumor was hyperdiploid with a non-amplified *MYCN*. He received local radiotherapy (450 cGy) followed by chemotherapy as per Pediatric Oncology Group study 9243: cyclophosphamide (7,350 mg/m²), doxorubicin (245 mg/m²), cisplatin (180 mg/m²), and etoposide (400 mg/m²). He went into complete remission and was well until his most current presentation. He was thin with pallor. No abdominal masses were palpated. Laboratory studies revealed iron deficiency anemia and an elevated ESR. An initial computed tomography (CT) scan

the area of the distal ileum that was previously suspected to be an unopacified loop of bowel (Fig. 1B). The mass was completely resected with negative margins (Fig. 1C). Histologically, the soft tissue mass included sheets of round cells having focal clear cell and rhabdoid features (Fig. 1D). FISH was positive for the EWS gene rearrangement confirming the diagnosis of GI CCS.

Rubino et al. [5] analyzed data on 544 childhood survivors of neuroblastoma and found 13 SMNs. The median age of neuroblastoma diagnosis in these patients was 17 months (range, 3–35 months). Further, the median time from diagnosis of neuroblastoma and occurrence of an SMN was 19.5 years (range, 5–37 years) which is consistent with the timing of presentation in our patient (14 years). The most common SMNs included thyroid (n = 5), breast (n = 3), and leukemia (n = 2). All but one patient received radiotherapy while only three received chemotherapy. Moreover, five of those receiving radiotherapy developed an SMN within the radiation field. This would suggest that radiotherapy may be an important risk factor in the development of SMNs in neuroblastoma patients. The proximity of our



**Fig. 1.** Initial FDG-PET image (**A**) demonstrates hypermetabolism within the right lower quadrant that corresponds to the distal ileum on the CT scan (**B**). The resected ileum mass (**C**) demonstrates clear cells on histopathology examination (**D**). [Color figure can be seen in the online version of this article, available at http://wileyonlinelibrary.com/journal/pbc]

was read as non-diagnostic. However an <sup>18</sup>F-fluoro-2-deoxy D-glucose positron emission tomography (FDG-PET) scan revealed an avid area of intensity in the presacral region (Fig. 1A). When the CT scan was retrospectively reviewed, this area of FDG-PET avidity corresponded to a 2.5 cm soft tissue intraluminal mass in

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patient's GI CCS primary to his previous site of radiation further supports this assumption. Although the significance of chemotherapy, particularly etoposide and cyclophosphamide, combined with radiotherapy in our patient and its influence on the development of his SMN is difficult to determine, all three patients that experienced an SMN in the Rubino et al. study received DNA topoisomerase II inhibitors.

The development of a SMN many years after neuroblastoma stresses the importance of long-term surveillance in childhood cancer survivors and highlights the potential impact therapeutic decisions have on our youngest and most vulnerable patients.

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